

# Middle ear gas exchange in isobaric counterdiffusion

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DUEKER, C. W., C. J. LAMBERTSEN, J. J. ROSOWSKI, AND J. C. SAUNDERS. *Middle ear gas exchange in isobaric counterdiffusion*. *J. Appl. Physiol.: Respirat. Environ. Exercise Physiol.* 47(6): 1239-1244, 1979.—Nitrous oxide entry into the middle ear gas space was studied in cats in relation to anesthesia and the vestibular dysfunction caused by isobaric inert gas counterdiffusion in diving. A catheter implanted in the auditory bulla was used for direct gas sampling and pressure measurements. Experiments were designed to evaluate the participation of the eustachian tube, mucosal blood vessels, and tympanic membrane in middle ear gas exchange. The eustachian tube did not contribute to N<sub>2</sub>O entry and the mucosal blood supply only contributed about one-third of the total N<sub>2</sub>O accumulation. Diffusion across the tympanic membrane accounted for most of the N<sub>2</sub>O entering the middle ear from ambient and respiratory environments containing N<sub>2</sub>O.

inert gas uptake; nitrous oxide; anesthesia; diving

THIS STUDY OF MIDDLE EAR gas exchange was prompted by the occurrence of incapacitating vestibular dysfunction in men breathing nitrogen-helium-oxygen or neon-helium-oxygen mixtures while surrounded by helium (5, 8, 9). These symptoms have now been identified, along with dermal gas lesions (9) and continuous venous gas embolization (7), as part of the isobaric counterdiffusion phenomenon, which is particularly important as a hazard in manned undersea activity. The process of inert gas counterdiffusion, though without such severe consequences, also occurs in gas inhalation anesthesia.

Whether vestibular dysfunction induced by counterdiffusion is caused by gas flux across the membrane of the round window, resulting in damaging gas phase development in the fluid of the inner ear (9), is not known. It is conceivable that the dysfunction may also be caused by gas embolization produced by transcutaneous isobaric counterdiffusion (7, 9). Middle ear gas exchange is also a relevant factor in general anesthesia for open ear surgery because nitrous oxide may cause elevated middle ear pressure (7, 11, 16).

Gas exchange in the middle ear can occur in three ways: gas can diffuse across the tympanic membrane, enter or leave via the eustachian tube, or diffuse between mucosal blood vessels and the middle ear cavity. Ingelstedt and Jonson (6) reported that middle ear ventilation depends primarily on eustachian tube function. More recently, Elner (4) used a technique similar to that used by Ingelstedt and Jonson to evaluate gas absorption from the middle ear indirectly.

Studies of gas transfer across the tympanic membrane in vitro (3) and with radioactive xenon (16) have suggested that the tympanic membrane, in spite of its extremely thin character (10), contributes very little to middle ear gas exchange. These studies, however, used indirect techniques or in vitro preparations. None of them was concerned with the exchange of gas after a shift in the respired and/or ambient gases.

The present experiments were designed to study the dynamics of gas exchange in the middle ear of living animals by means of direct sampling techniques. Three different experimental designs permitted determination of the respective contributions of the eustachian tube, tympanic membrane, and mucosal blood vessels to middle ear gas exchange.

## GENERAL METHODS

Adult male cats were used. The auditory bulla of the cat provides a convenient site for gas sampling and connects directly with the middle ear space. After induction of anesthesia with ketamine (avg dose 20 mg/kg), tracheal intubation was performed via the mouth. Ketamine provided maintenance anesthesia as needed; incremental doses up to a maximum of 60 mg/kg were given. The bulla was exposed by using a ventral approach modified from that of Paparella and Hohmann (13). After drilling a hole into the bulla, a 16-gauge polyethylene vascular catheter was inserted into the bulla. An attached plastic flange fixed the catheter tip 2 mm into the bulla cavity. The flange was attached to the bulla with fast-drying dental acrylic cement; the flange and exposed bulla were then covered with acrylic filler and the incision was closed in layers. To facilitate visualization of the tympanic membrane, a 1-cm incision was made in the tragus; each side of this incision was maintained with wire suture. The cat was then allowed to recover from surgery.

Nitrous oxide was used as the experimental gas because its effects on middle ear pressure have been reported (11, 15, 18).

The experiments were performed either in air or in a glove box filled with N<sub>2</sub>O and oxygen. A pump circuit provided temperature and humidity control for the glove box. Carbon dioxide expired by the animals was removed by alkali absorbent. An infrared gas analyzer (Beckman) measured N<sub>2</sub>O concentration for continuous recording. Oxygen partial pressure was measured by a paramagnetic oxygen analyzer (Servomex).

Before the experiment, each cat was anesthetized. Air

was injected into the bulla catheter, and the motion of the tympanic membrane was then observed to verify system tightness. A low-dead-space three-way stopcock was then attached to the bulla catheter. Continuous middle ear pressure determinations were made with a water-filled strain gauge transducer (Statham) and recorded.

Gas samples were withdrawn from the catheter into 100- $\mu$ l gas-tight syringes (Precision Sampling Pressure-Lok series A-2) at about 1, 3, 6, 9, 12, and 15 h after the experiment began. An initial sample of 100  $\mu$ l was withdrawn to remove the dead-space gas trapped in the catheter, and a second sample was then drawn for analysis. The first sample was then reinjected into the bulla to minimize ear volume changes. Each sample was analyzed for CO<sub>2</sub>, N<sub>2</sub>O, O<sub>2</sub>, and N<sub>2</sub> with a gas chromatograph (Varian Aerograph 1420). Peak areas of the chromatogram were digitally integrated (Hewlett-Packard 3700A) and the resulting values were converted to gas percent volumes.

To minimize the effect of slight changes in inspired or ambient N<sub>2</sub>O, bulla N<sub>2</sub>O concentration was expressed as a fraction of the inspired N<sub>2</sub>O or the ambient N<sub>2</sub>O. The mean inspired or ambient concentration over the hour preceding each bulla sample was used in calculating this ratio.

Three experimental situations were studied: *I*, cats breathing N<sub>2</sub>O-O<sub>2</sub> while surrounded by N<sub>2</sub>O-O<sub>2</sub>; *II*, cats breathing N<sub>2</sub>O-O<sub>2</sub> while surrounded by air; and *III*, cats breathing air while surrounded by N<sub>2</sub>O.

#### Situation I. N<sub>2</sub>O Breathing in N<sub>2</sub>O Atmosphere

**Procedures.** This situation was designed to measure maximal entry of gas into the middle ear. Seven cats

breathed 80% N<sub>2</sub>O-20% O<sub>2</sub> while surrounded by 75% N<sub>2</sub>O-25% O<sub>2</sub>.

Each cat was studied in a closed plastic box large enough to accommodate two cats. Rubber gloves mounted in the side of the box allowed access to the animals. The animals breathed spontaneously. Anesthesia, gas sampling, and pressure measurements were performed in accordance with procedures described in GENERAL METHODS.

**Results.** Figure 1A illustrates the course of N<sub>2</sub>O entry into the middle ear for five of these animals. After a rapid rise to about 80% of inspired N<sub>2</sub>O within the 1st h, the rate of increase slowed with time and reached a plateau of 90% of the inspired gas at 6 h. A long balloon catheter was placed in the nose of one cat (Fig. 1A, *expt 7*) and inflated with water to obstruct the orifice of the eustachian tube almost completely. The extent of the eustachian tube seal was verified by injecting air into the bulla and showing by the pressure traces that the eustachian tube could not completely vent the increased middle ear pressure. Gas entry in this cat did not differ detectably from that in the cats with patent eustachian tubes (Fig. 1A). To assess the effect of sampling per se on gas equilibrium, gas was not sampled in two cats until 8 h after the experiment began, at which time equilibrium between middle ear and ambient N<sub>2</sub>O had been achieved.

Figure 1B shows sequences from a typical middle ear pressure tracing from one animal in *situation I*. During the period of rapid N<sub>2</sub>O entry, pressure rose by as much as 9 cmH<sub>2</sub>O in some animals, and then fell to ambient. These decreases in pressure probably accompanied eustachian tube opening because pressure tracings from the cat with the obstructed eustachian tube did not show the marked changes seen in animals with patent tubes. Maximum bulla pressure in *situation I* slowly decreased over

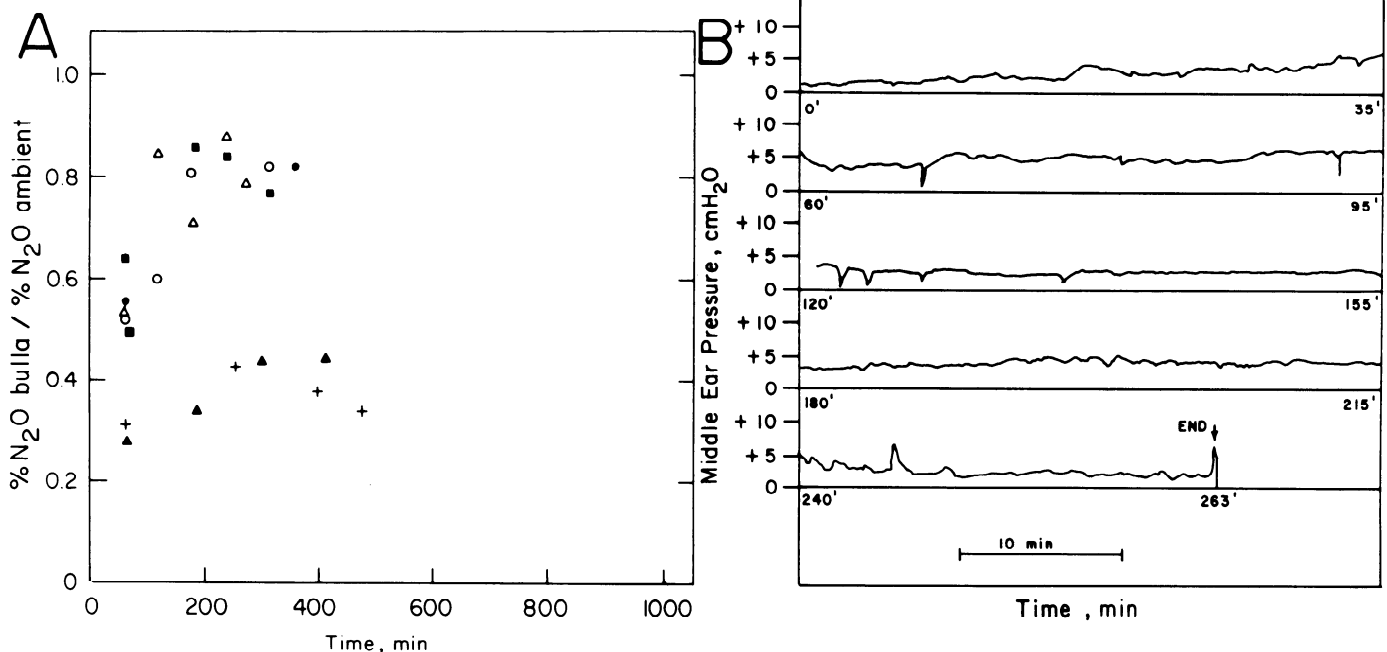


FIG. 1. N<sub>2</sub>O breathing-N<sub>2</sub>O ambient situation (*I*). A: attainment of near-equilibrium between middle ear and inspired N<sub>2</sub>O concentrations; results represent essentially exponential increases from initial middle ear N<sub>2</sub>O value of zero. Five experiments: 1, ●; 2, +; 3, ▲; 4, ■; 7, ○. B:

5 traces, each lasting 35 min showing time course of pressure changes in middle ear during approximately 10 h of exposure. *Rise*: entry of gas; *abrupt fall*: venting via eustachian tube (*expt 1*).

time and returned to a control level within 8 h after initiation of the experimental procedure.

### Situation II. N<sub>2</sub>O Breathing in Air Atmosphere

Results from *situation I* demonstrated a rapid rise of N<sub>2</sub>O concentration in the bulla cavity when the eustachian tube was both blocked and unblocked, indicating that the eustachian tube does not contribute to this uptake. *Situation II* was designed to study the contribution of a single pathway, the mucosal blood vessels, to middle ear gas exchange.

**Procedure.** Five cats were anesthetized according to procedures outlined in GENERAL METHODS. Four were prepared by endotracheal intubation via the mouth; one of these four (*expt 12*) also had its eustachian tube blocked. The cuff of the tube was inflated until positive-pressure ventilation resulted in no leakage around the tube. One cat (*expt 11*) inhaled N<sub>2</sub>O through an oronasal breathing mask, rather than through an endotracheal tube.

The cats spontaneously breathed 80% N<sub>2</sub>O-20% O<sub>2</sub> supplied through an unvalved nonrebreathing system at a flow rate of 1 l/min. Ambient gas was room air. Respiratory N<sub>2</sub>O and O<sub>2</sub> concentrations were measured continuously, as described in GENERAL METHODS, with a catheter placed in the endotracheal tube connector or under the mask. Middle ear pressure measurements and gas sampling also followed the procedures described in GENERAL METHODS.

**Results.** Figure 2A shows N<sub>2</sub>O in the bulla as a ratio of inspired N<sub>2</sub>O in the five cats. In the 1st h, this ratio reached 20%; it leveled off at about 35% after 5 h. The cat that breathed through the mask (*Fig. 2A, expt 11*) had a gas entry equivalent to that of the intubated cats.

If retrograde gas flow from nasopharynx to middle ear contributed to N<sub>2</sub>O exchange, the masked cat (nasopharynx exposed to respired N<sub>2</sub>O) would have shown a faster rise in middle ear N<sub>2</sub>O. This was not the case, and to evaluate eustachian tube retrograde flow further, a nasal balloon was placed in one of the intubated cats (eustachian tube exposed to ambient air) as in *situation I* (*Fig. 2A, expt 12*). This procedure did not affect N<sub>2</sub>O entry into the middle ear.

Figure 2B shows a representative pressure tracing from one animal in this experimental situation. Pressure elevations of as much as 20 cmH<sub>2</sub>O began promptly with N<sub>2</sub>O breathing, and were followed by eustachian tube venting. The pressure shifts continued and showed only a slight decrease over time. This was in marked contrast to the pressure tracings of animals both breathing and surrounded by N<sub>2</sub>O (*situation I*) (*Fig. 1B*). Blockage of the eustachian tube in one cat resulted in a persistent positive pressure without cyclic rapid pressure falls attributable to eustachian tube venting.

### Situation III. Air Breathing in N<sub>2</sub>O Atmosphere

Breathing N<sub>2</sub>O while surrounded by air did not account for the full entry of N<sub>2</sub>O shown in the total (respiratory and ambient) N<sub>2</sub>O exposure of *situation I*. This third study was therefore conducted to determine the contribution to gas entry into the bulla by diffusion through the tympanic membrane itself.

**Procedure.** Six cats were anesthetized and intubated. After intubation, they were placed in the glove box and spontaneously breathed air delivered from outside the box through an unvalved nonrebreathing apparatus. The exhaust from the breathing bag was vented outside the box. A sampling line in the endotracheal tube connector

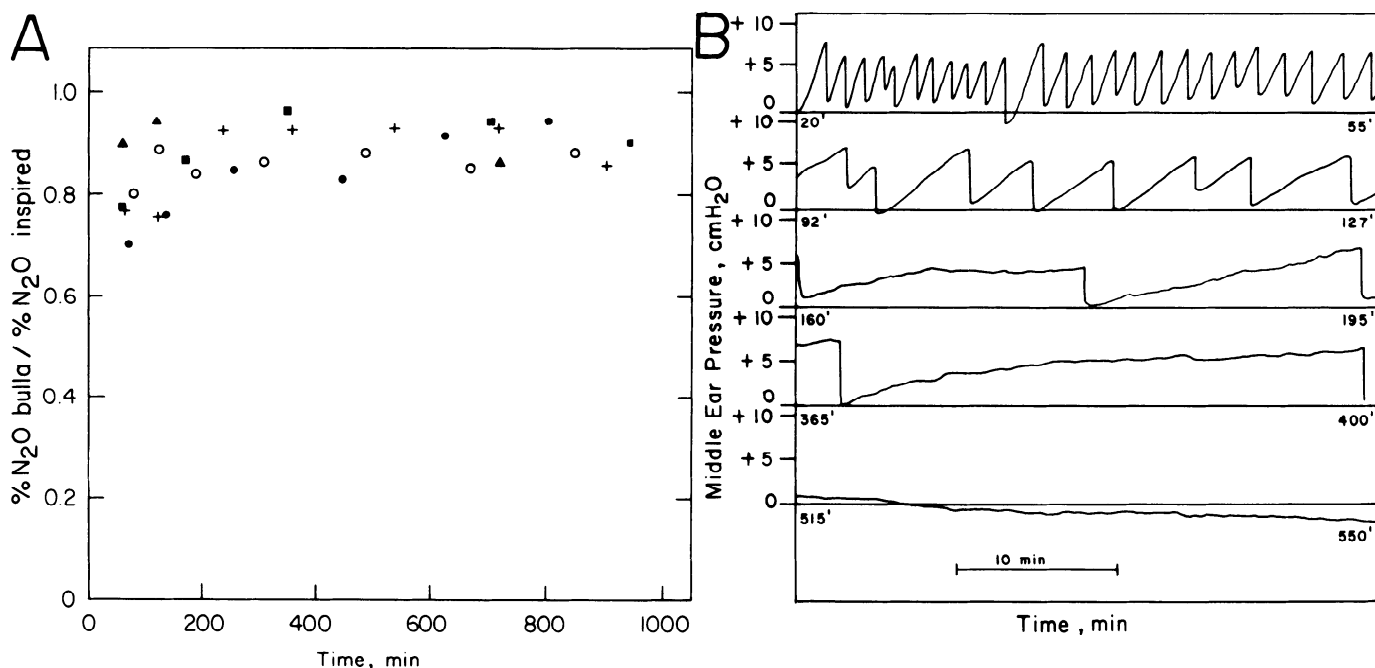


FIG. 2. N<sub>2</sub>O breathing-air ambient situation (II). A: relatively low equilibrium values for composite effects of N<sub>2</sub>O entry (via respiratory gas) and N<sub>2</sub>O loss (via tympanic membrane). Five experiments: 8, ●; 9, +; 10, ▲; 11, ○; 12, ■. B: representative time course of middle ear

pressure changes (*expt 8*). Repeated rises and falls indicate continuous entry of N<sub>2</sub>O throughout experiment despite low stable-state N<sub>2</sub>O concentrations shown in A.

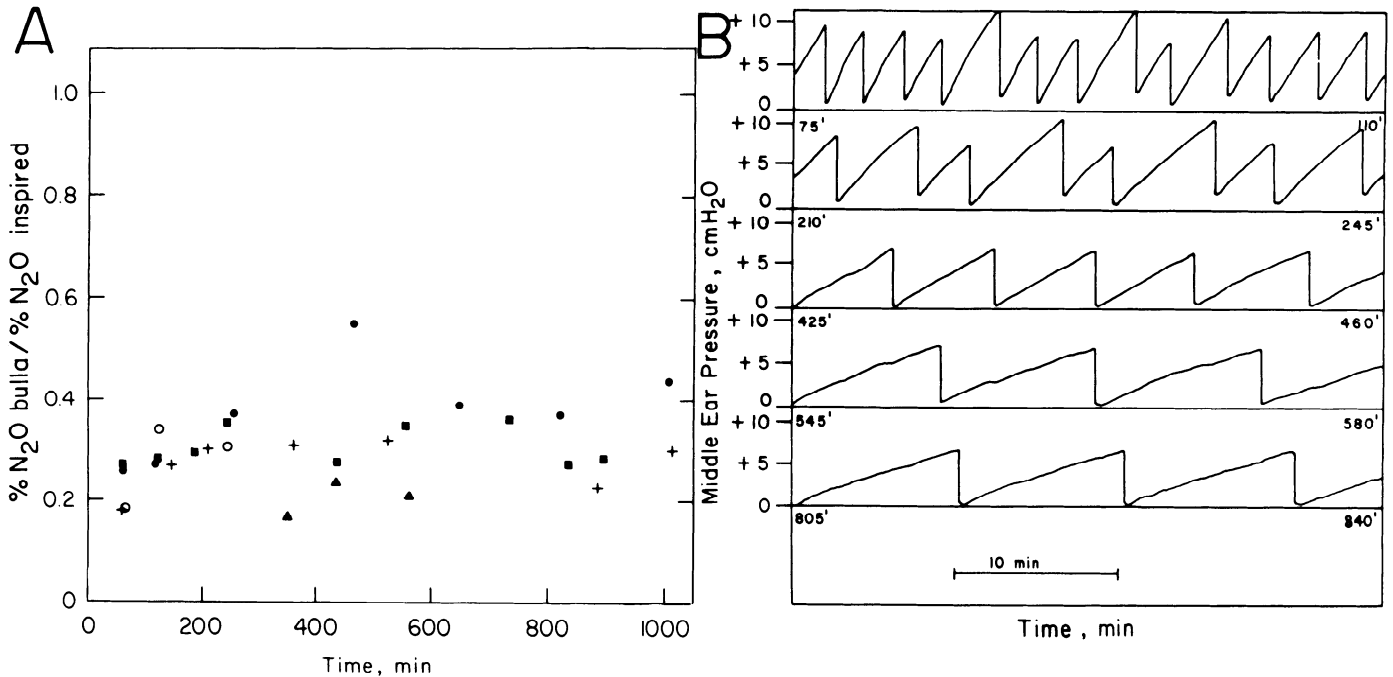


FIG. 3. Air breathing- $N_2O$  ambient situation (III). A: rise in ratio from initial zero level indicating entry of  $N_2O$  via tympanic membrane. Six experiments: 13, +; 14,  $\blacktriangle$ ; 15,  $\circ$ ; 16,  $\bullet$ ; 17,  $\triangle$ ; 18,  $\blacksquare$ . B: tracing from

representative experiment (no. 15) demonstrating absence of the pressure increases characteristic of  $N_2O$  breathing situations (see Figs. 1B and 2B).

was used to determine respired gas composition. As the cats breathed air, they were surrounded by a circulated atmosphere of 75%  $N_2O$ -25%  $O_2$ . Pressure measurements and gas sampling followed the methods described.

One cat was surrounded, except for his head, with a plastic bag flushed with air, to eliminate the possibility that some  $N_2O$  entry into the middle ear might have resulted from gas entering through the skin (1). Two cats had nasal balloons placed (as in situations I and II) to obstruct the eustachian tubes.

**Results.** Figure 3A shows middle ear  $N_2O$  concentration changes in cats breathing air while surrounded by  $N_2O$ . In this circumstance the ratio is expressed as bulla concentration/ambient  $N_2O$ . This experiment showed the widest range of  $N_2O$  concentrations. At 1 h, the ratios ranged between 30% and 65%; at 4 h, the range was 40–85%. The cat with no cutaneous exposure to  $N_2O$  also showed middle ear  $N_2O$  entry that fell within these ranges. Eustachian tube blockage did not affect  $N_2O$  level. Figure 3B shows the absence of the pressure fluctuations (from *expt 15*) characteristic of  $N_2O$  breathing situations.

## DISCUSSION

During ordinary respiration of  $N_2O$ , all parts of the body give up  $N_2$  as they take up  $N_2O$ . Diffusion rates into a gas-filled space like the middle ear depend on pressure gradients, area of diffusion surface, and length of diffusion path, as well as gas molecular weight and solubility. On the basis of gas permeabilities,  $N_2O$  should enter the middle ear space, by several pathways, 30 times faster than  $N_2$  can leave.

During anesthesia with  $N_2O$ , the relatively large amount of  $N_2O$  entering closed gas spaces can cause

dangerous increases in volume (distensible spaces) or pressure (rigid spaces). This has been reported in the middle ear (11, 15, 18) and the cranial vault (17), and in cases of intestinal obstruction (2), pneumothorax (2), and venous air embolism (12).

In semirigid spaces such as those of the middle ear, the imbalance between gas entry and loss causes a rise in pressure. The eustachian tube then opens to relieve pressure, venting the excess gas. The amount of gas vented approximates the net gas entry, that is, the difference between the amount of  $N_2O$  entering and the amount of  $N_2$  leaving. If the volume of the middle ear is known, Boyle's law can be used to calculate the amount of gas exhausted at each eustachian tube opening.

Assuming a middle ear volume in the cat of 2 ml, cats in the  $N_2O$  breathing- $N_2O$  ambient situation (I) had an uptake of 0.0088 ml/min in the 1st h, with a decrease to 0.0024 ml/min during the 2nd h. The entry rate of  $N_2O$  decreased as the middle ear approached equilibrium with the surrounding  $N_2O$ , lowering the pressure gradient for diffusion. As seen in Fig. 1B, middle ear pressure was equal to atmospheric pressure at the time corresponding to maximum middle ear  $N_2O$  concentration.

Cats breathing  $N_2O$  while surrounded by air (II) never achieved equilibrium between respiratory gas and the middle ear space (Fig. 2B). The initial rate of gas uptake, 0.0054 ml/min, was slower than that of situation I (0.0088 ml/min), but even after 9 h  $N_2O$  still entered at 0.0014 ml/min.

### Role of Eustachian Tube

The results of this study do not support current concepts of middle ear gas exchange, which indicate that when a cat is surrounded by  $N_2O$ , flow up the eustachian

tube should hasten the increase of middle ear N<sub>2</sub>O concentration (4, 6). Almost complete blockage of eustachian tubes under this condition failed to modify N<sub>2</sub>O accumulation. It was considered possible that during endotracheal N<sub>2</sub>O breathing while surrounded by air, the eustachian tube might allow air to enter the middle ear, diluting the N<sub>2</sub>O. This was unlikely, considering the positive-pressure gradient from middle ear to nasopharynx. That it actually did not occur was shown by obstructing the eustachian tube almost completely. Similarly, breathing N<sub>2</sub>O by oronasal mask would increase middle ear N<sub>2</sub>O entry if N<sub>2</sub>O could flow up the eustachian tube. However, the amount of entry was the same for the oronasal mask as for the endotracheal N<sub>2</sub>O inhalation.

Ingelstedt and Jonson (6) calculated eustachian tube ventilation by measuring ear volume changes after inducing negative pressure within the middle ear. Elner (4) used similar calculations to estimate the amount of gas entering by way of the eustachian tube to replace the volume of gas absorbed from the middle ear into the mucosal vessels. In neither of these studies was eustachian tube function measured under conditions of variation in inhaled or ambient gas.

### Role of Tympanic Membrane

Previous studies have concluded that the tympanic membrane contributes little to middle ear gas exchange. Elner (3) exposed cadaver tympanic membrane to CO<sub>2</sub> in a diffusion chamber; extrapolation of these results to O<sub>2</sub> and N<sub>2</sub> led to the report that diffusion across the tympanic membrane represented only 0.5–1.0% of the amount of gas venting through the eustachian tube. Riu et al. (16) has stated that transfer of radioactive xenon through the tympanic membrane was nonexistent or insignificant.

During N<sub>2</sub>O breathing while surrounded by air (II), N<sub>2</sub>O reaching the middle ear space arrived via the mucosal and tympanic membrane blood vessels. After 12 h, the blood concentration of N<sub>2</sub>O should have been equivalent to that of the inspired mixture (80% N<sub>2</sub>O). However, middle ear N<sub>2</sub>O did not rise above 35%. Outward flow of gas through the eustachian tube did not alter the composition of middle ear gas, and there was no inward passage. Therefore, the failure in N<sub>2</sub>O equilibrium was most probably caused by outward diffusion of N<sub>2</sub>O through the tympanic membrane, accompanied by inward diffusion of nitrogen into the middle ear space through the tympanic membrane.

To measure the diffusion of N<sub>2</sub>O across the tympanic membrane, air-breathing cats surrounded by N<sub>2</sub>O were studied (*situation III*). The only available routes for N<sub>2</sub>O entry were through the skin, around the endotracheal tube, up the eustachian tube, and across the tympanic membrane. Blocking the cutaneous route did not affect entry rate. Measurement of N<sub>2</sub>O from all sources in the respiratory gas of the endotracheal tube showed that it was consistently less than 4%. The eustachian tube did not serve as a source of N<sub>2</sub>O. Thus, N<sub>2</sub>O reaching the middle ear in the air breathing-N<sub>2</sub>O ambient situation (*III*) could only have come through the tympanic membrane.

The significant role played by diffusion across the tympanic membrane has been demonstrated in two ways. Outward diffusion markedly limited the extent of N<sub>2</sub>O accumulation derived from mucosal blood supply, and inward diffusion resulted in a rapid increase of N<sub>2</sub>O. Presumably, without loss of N<sub>2</sub>O to the capillaries of the middle ear, there would be prompt gas equilibration between atmosphere and the middle ear of cats surrounded by N<sub>2</sub>O.

### Relation to Other Isobaric Counterdiffusion States

The aim of this study of nitrous oxide exchange in the middle ear cavity was to increase understanding of the process of isobaric inert gas diffusion among atmosphere, middle ear, and inner ear. The results indicate that nitrous oxide passes through the tympanic membrane more rapidly than had previously been believed. It is therefore likely that at high ambient pressures, very rapid movement of helium or other gases across the tympanic membrane occurs. An extension of the present study has led to a detailed mathematical model of middle ear gas exchange (14). Direct measurement of transtympanic diffusion of helium, nitrogen, and other respirable gases used at increased ambient pressure is necessary to relate these findings to the severe vestibular disruption associated with isobaric counterdiffusion in man.

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### REFERENCES

1. CULLEN, B. F., AND E. I. EGER II. Diffusion of nitrous oxide, cyclopropane, and halothane through human skin and amniotic membrane. *Anesthesiology* 36: 168–173, 1972.
2. EGER, E. I., II, AND F. J. SAIDMAN. Hazards of nitrous oxide anesthesia in bowel obstruction and pneumothorax. *Anesthesiology* 26: 61–66, 1965.
3. ELNER, Å. Gas diffusion through the tympanic membrane. *Acta Oto-Laryngol.* 69: 185–191, 1970.
4. Elner, Å. Indirect determination of gas absorption from the middle ear. *Acta Oto-Laryngol.* 74: 191–196, 1972.
5. GRAVES, D. J., J. IDICULA, C. J. LAMBERTSEN, AND J. A. QUINN. Bubble formation resulting from counterdiffusion supersaturation: a possible explanation for isobaric inert gas 'urticaria' and vertigo. *Phys. Med. Biol.* 18: 256–264, 1973.
6. INGELSTEDT, S., AND B. JONSON. Mechanisms of gas exchange in the normal human middle ear. *Acta Oto-Laryngol. Suppl.* 224: 452–461, 1966.
7. LAMBERTSEN, C. J., J. P. W. CUNNINGTON, AND J. R. M. COWLEY. The dynamics and composition of spontaneous, continuous gas embolism in the pig during isobaric gas counterdiffusion (Abstract). *Federation Proc.* 34: 452, 1975.
8. LAMBERTSEN, C. J., R. GELFAND, R. E. PETERSON, R. STRAUSS, W. B. WRIGHT, J. G. DICKSON, JR., C. PUGLIA, AND R. W. HAMILTON, JR. Human tolerance to He, Ne, and N<sub>2</sub> at respiratory gas

- densities equivalent to He-O<sub>2</sub> breathing at depths of 1200, 2000, 3000, 4000, and 5000 feet of sea water. *Aviat. Space Environ. Med.* 48: 843-855, 1977.
9. LAMBERTSEN, C. J., AND J. IDICULA. A new gas lesion syndrome in man induced by isobaric gas counterdiffusion. *J. Appl. Physiol.* 39: 434-443, 1975.
  10. LIM, D. J. Tympanic membrane. Electron microscopic observation. I. Pars tensa. *Acta Oto-Laryngol.* 66: 181-198, 1968.
  11. MATZ, G. J., C. G. RATTENBORG, AND D. A. HOLADAY. Effects of nitrous oxide on middle ear pressure. *Anesthesiology* 28: 948-950, 1967.
  12. MUNSON, E. S., AND H. C. MERRICK. Effect of nitrous oxide on venous air embolism. *Anesthesiology* 27: 783-787, 1966.
  13. PAPARELLA, M. M., AND A. HOHMANN. Surgical techniques for otological and auditory research. *Ann. Otol. Rhinol. Laryngol.* 71: 203-212, 1962.
  14. RANADE, A. *Analysis of Inert Gas Exchange in the Middle Ear* (PhD thesis). Philadelphia, PA: Univ. of Pennsylvania, 1977.
  15. RASMUSSEN, P. E. Middle ear and maxillary sinus during nitrous oxide anesthesia. *Acta Oto-Laryngol.* 63: 7-16, 1967.
  16. RIU, R., L. FLOTTE, J. BOUCHE, AND R. LE DEN. *La physiologie de la Trompe d'Eustache*. Paris: Librairie Arnette, 1966.
  17. SAIDMAN, L. J., AND E. I. EGER II. Changes in cerebrospinal fluid pressure during pneumoencephalography under nitrous oxide anesthesia. *Anesthesiology* 26: 67-72, 1965.
  18. THOMSEN, K. A., K. TERKILDSEN, AND I. ARNFRED. Middle ear pressure variations during anesthesia. *Arch. Otolaryngol.* 82: 609-611, 1965.

